

# African Journal of Pharmacy and Pharmacology

Volume 10 Number 32, 29 August, 2016  
ISSN 1996-0816



## ABOUT AJPP

The **African Journal of Pharmacy and Pharmacology (AJPP)** is published weekly (one volume per year) by Academic Journals.

**African Journal of Pharmacy and Pharmacology (AJPP)** is an open access journal that provides rapid publication (weekly) of articles in all areas of Pharmaceutical Science such as Pharmaceutical Microbiology, Pharmaceutical Raw Material Science, Formulations, Molecular modeling, Health sector Reforms, Drug Delivery, Pharmacokinetics and Pharmacodynamics, Pharmacognosy, Social and Administrative Pharmacy, Pharmaceutics and Pharmaceutical Microbiology, Herbal Medicines research, Pharmaceutical Raw Materials development/utilization, Novel drug delivery systems, Polymer/Cosmetic Science, Food/Drug Interaction, Herbal drugs evaluation, Physical Pharmaceutics, Medication management, Cosmetic Science, pharmaceuticals, pharmacology, pharmaceutical research etc. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in AJPP are peer-reviewed.

### Contact Us

Editorial Office: [ajpp@academicjournals.org](mailto:ajpp@academicjournals.org)

Help Desk: [helpdesk@academicjournals.org](mailto:helpdesk@academicjournals.org)

Website: <http://www.academicjournals.org/journal/AJPP>

Submit manuscript online <http://ms.academicjournals.me/>

## Editors

### **Himanshu Gupta**

*Department of Pharmacy Practice  
University of Toledo  
Toledo, OH  
USA.*

### **Prof. Zhe-Sheng Chen**

*College of Pharmacy and Health Sciences  
St. John's University  
New York,  
USA.*

### **Dr. Huma Ikram**

*Neurochemistry and Biochemical  
Neuropharmacology Research Unit,  
Department of Biochemistry,  
University of Karachi  
Karachi-75270  
Pakistan*

### **Dr. Shreesh Kumar Ojha**

*Molecular Cardiovascular Research Program  
College of Medicine  
Arizona Health Sciences Center  
University of Arizona  
Arizona,  
USA.*

### **Dr. Vitor Engracia Valenti**

*Departamento de Fonoaudiologia  
Faculdade de Filosofia e Ciências,  
UNESP  
Brazil.*

### **Dr. Caroline Wagner**

*Universidade Federal do Pampa  
Avenida Pedro Anunciação  
Brazil.*

## Associate Editors

### **Dr. B. Ravishankar**

*SDM Centre for Ayurveda and Allied Sciences,  
SDM College of Ayurveda Campus,  
Karnataka  
India.*

### **Dr. Natchimuthu Karmegam**

*Department of Botany,  
Government Arts College,  
Tamil Nadu,  
India.*

### **Dr. Manal Moustafa Zaki**

*Department of Veterinary Hygiene and  
Management  
Faculty of Veterinary Medicine,  
Cairo University  
Giza,  
Egypt.*

### **Prof. George G. Nomikos**

*Takeda Global Research & Development Center  
USA.*

### **Prof. Mahmoud Mohamed El-Mas**

*Department of Pharmacology,  
Faculty of Pharmacy  
University of Alexandria,  
Alexandria,  
Egypt.*

### **Dr. Kiran K. Akula**

*Electrophysiology & Neuropharmacology Research  
Unit  
Department of Biology & Biochemistry  
University of Houston  
Houston, TX  
USA.*

## Editorial Board

**Prof. Fen Jicai**

*School of life science, Xinjiang University, China.*

**Dr. Ana Laura Nicoletti Carvalho**

*Av. Dr. Arnaldo, 455, São Paulo, SP. Brazil.*

**Dr. Ming-hui Zhao**

*Professor of Medicine  
Director of Renal Division, Department of Medicine  
Peking University First Hospital  
Beijing 100034  
PR. China.*

**Prof. Ji Junjun**

*Guangdong Cardiovascular Institute, Guangdong General Hospital, Guangdong Academy of Medical Sciences, China.*

**Prof. Yan Zhang**

*Faculty of Engineering and Applied Science,  
Memorial University of Newfoundland,  
Canada.*

**Dr. Naoufel Madani**

*Medical Intensive Care Unit  
University hospital Ibn Sina, Univesity Mohamed V  
Souissi, Rabat,  
Morocco.*

**Dr. Dong Hui**

*Department of Gynaecology and Obstetrics, the 1st hospital, NanFang University, China.*

**Prof. Ma Hui**

*School of Medicine, Lanzhou University, China.*

**Prof. Gu HuiJun**

*School of Medicine, Taizhou university, China.*

**Dr. Chan Kim Wei**

*Research Officer  
Laboratory of Molecular Biomedicine,  
Institute of Bioscience, Universiti Putra,  
Malaysia.*

**Dr. Fen Cun**

*Professor, Department of Pharmacology, Xinjiang University, China.*

**Dr. Sirajunnisa Razack**

*Department of Chemical Engineering, Annamalai University, Annamalai Nagar, Tamilnadu, India.*

**Prof. Ehab S. EL Desoky**

*Professor of pharmacology, Faculty of Medicine Assiut University, Assiut, Egypt.*

**Dr. Yakisich, J. Sebastian**

*Assistant Professor, Department of Clinical Neuroscience R54  
Karolinska University Hospital, Huddinge  
141 86 Stockholm ,  
Sweden.*

**Prof. Dr. Andrei N. Tchernitchin**

*Head, Laboratory of Experimental Endocrinology and Environmental Pathology LEEPA  
University of Chile Medical School,  
Chile.*

**Dr. Sirajunnisa Razack**

*Department of Chemical Engineering,  
Annamalai University, Annamalai Nagar, Tamilnadu, India.*

**Dr. Yasar Tatar**

*Marmara University,  
Turkey.*

**Dr Nafisa Hassan Ali**

*Assistant Professor, Dow institute of medical technology  
Dow University of Health Sciences, Chand bbi Road, Karachi,  
Pakistan.*

**Dr. Krishnan Namboori P. K.**

*Computational Chemistry Group, Computational Engineering and Networking,  
Amrita Vishwa Vidyapeetham, Amritanagar, Coimbatore-641 112  
India.*

**Prof. Osman Ghani**

*University of Sargodha,  
Pakistan.*

**Dr. Liu Xiaoji**

*School of Medicine, Shihezi University,  
China.*

**ARTICLES**

- Study of antidiarrheal and anthelmintic activity methanol extract of *Commelina benghalensis* leaves** 657  
Mohammad Shah Hafez Kabir, Abul Hasanat, Tanvir Ahmad Chowdhury,  
Mohammad Mamun Ur Rashid, Mohammed Munawar Hossain and  
Shabbir Ahmed
- Potential impact of urocortin I on sperm count, motility and sex hormone profiles in normal adult rats** 665  
Hany A. El kattawy, Mahmoud A. Naga, Ahmed Shata and Mohamed A. Elbaky

## Full Length Research Paper

## Study of antidiarrheal and anthelmintic activity methanol extract of *Commelina benghalensis* leaves

Mohammad Shah Hafez Kabir<sup>1</sup>, Abul Hasanat<sup>1</sup>, Tanvir Ahmad Chowdhury<sup>1</sup>, Mohammad Mamun Ur Rashid<sup>2\*</sup>, Mohammed Munawar Hossain<sup>1</sup> and Shabbir Ahmed<sup>1</sup>

<sup>1</sup>Department of Pharmacy, International Islamic University Chittagong, Bangladesh.

<sup>2</sup>Department of Pharmaceutical Sciences, North South University, Dhaka, Bangladesh.

Received 21 August 2015; Accepted 14 October, 2015

The objective of this study was to examine the restorative effects of methanol extract of *Commelina benghalensis* leaves in antidiarrheal activity in Swiss albino mice model and anthelmintic activity in *Tubifex tubifex*. Antidiarrheal activity study in Swiss albino mice were divided into four groups. Group I was treated as the control group and received 10 ml/kg of 2% Tween-80 orally; Group II served as a positive control and took standard drug (loperamide) in 5 mg/kg orally; Groups III and IV were the test groups which received the methanol extract of *C. benghalensis* orally at 200 and 400 mg/kg, respectively. Dependent upon the model comprehensive weight of dry feces, aggregate weight of wet defecation, length of intestinal travel and intestinal weight were gathered. At long last, information were investigated using restricted analysis of variance (ANOVA) took after by Dunnett test. Anthelmintic action on aquarium worm *T. tubifex* by utilizing three focuses of 10, 5, and 2.5 mg/ml of *C. benghalensis* movement was assessed by *in vitro* system. The property of the extract on defecation, intestinal transit and intestinal fluid accumulation (antienterpooling) were assessed in castor oil induced diarrhea. Leaves extract of *C. benghalensis* at 200 and 400 mg/kg exhibited significant ( $p < 0.05$ , 0.01 and 0.001) and dose dependent antidiarrheal probable in castor oil induced diarrhea. The diarrhea was inhibited by 27.59 and 55.17 at doses of 200 and 400 mg/kg in that order. The extract were found to have an antienterpooling in castor oil affected test Swiss albino mice's by decreasing both weight and volume of intestinal substance, and obviously reducing the intestinal travel similar to that of the standard medication, loperamide (5 mg/kg). Anthelmintic activity of methanol extract *C. benghalensis* show the time taken for paralysis (12.38, 16.25 and 22.17 min) and time taken for death (21.39, 45.64 and 57.25 min) at doses of 10, 5, and 2.5, respectively. The methanol extract of *C. benghalensis* demonstrated antidiarrheal effect in Swiss albino mice model and anthelmintic action on aquarium worm *T. tubifex*. In addition, the extract was discovered to be safe at measurements of 1000, 2000, 3000, and 4000 mg/kg in mice model. The discoveries propose the legitimacy of the much lauded impact of *C. benghalensis* as antidiarrheal and anthelmintic specialists in expected home grown medication.

**Key words:** *Commelina benghalensis*, anti-diarrhea, Swiss albino mice, anthelmintic, *Tubifex tubifex*.

### INTRODUCTION

Nature has been a recurrent source of pharmacologically dynamic atoms and curative herbs have been utilized by incalculable human eras (Zheng et al., 2015). *Commelina*

*benghalensis* Linn. (Commelinaceae) is a perennial herb native to tropical Asia and Africa, commonly known as Bengal day flower or Dew flower. It is a large, straggling

annual herb up to 40 cm long with rooting at basal nodes and characterized by attractive small bluish-violet flowers. Leaves are ovate-elliptic or oblong, shortly triangular, bright green in color and 4 to 7 cm long. The spathes are green, funnel-shaped, compressed and about 1.5 cm long. Capsules are broadly ovoid-oblong and 4 to 5 mm long (Zheng et al., 2015).

*C. benghalensis* is utilized as a part of customary medication outline to treat different diseases. It is used for the treatment of headache, constipation, leprosy, fever, snake bite and jaundice (Hasan et al., 2008; Yusuf et al., 1994; Kirtikar and Basu, 1980). It is also used in the treatment of mouth thrush (Ssenyonga and Brehony, 1993), insanity (Tabuti et al., 2003), epilepsy (Okello and Ssegawa, 2007) and psychosis (Adjanohoun, 1993). In Lesotho, it is applied to treat infertility in women and in India it is used as bitter, laxative, anti-inflammatory, demulcent, emollient and depressant (Jayvir et al., 2007). In China, it is used as diuretic and febrifuge (Hong and Deflipsis, 2000). In Pakistan it is used as vegetable (Qaiser and Jafri, 1975). In Nepal, the paste of the plant is utilized to treat smolders and juice of the roots is utilized to treat acid reflux (Manandhar and Sanjay, 2000).

In a past study, antimicrobial action of watery extract of *C. benghalensis* was assessed (Sharma and Sharma, 2010). Potential calming and anxiolytic exercises of diverse portions of the plant are accounted for in the writing (Hasan et al., 2009). The plant is also reported with remarkable antioxidant, antitumor and anticancer, and thrombolytic activity (Zheng et al., 2015; Mbazima et al., 2008; Rahman et al., 1999; Chowdhury et al., 2015). Protective activity of the roots extract against paracetamol induced hepatic damage in Wistar rats has been reported (Paresh and Chanda, 2008). Phytochemical investigations on *C. benghalensis* have revealed the existence of alkaloid, volatile oil, wax (Raju et al., 2007), vitamin C, vitamin A and  $\beta$ -carotene (Guerrant et al., 2001). Diarrhea can be characterized as a modification in the ordinary solid discharge, portrayed by a circumstance in which a grown-up every day stools surpasses 300 g and contains 60 to 95% water (Zavala et al., 1998). Looseness of the bowels is one of the primary drivers of newborn child passing particularly in underdeveloped nation (Galheigo et al., 2015). It represents more than 5 million passing in overall every year in newborn children and offspring of under 5 years. Therapeutic plants are potential wellsprings of antidiarrheal medications (Van et al., 2015; Chowdhury et al., 2015).

In Bangladesh, one third of the total child death burden is due to diarrhea (Patzl et al., 2015). Use of traditional medicines to combat the episodes of diarrhea has been emphasized by WHO in its Diarrhea Control Program

(Wansi et al., 2007). Medicinal plants have been reported for their efficiency in the treatment of diarrhea, thanks to the contribution of many researchers (Dooley et al., 2015). Gastrointestinal nematodes (GIN) cause genuine monetary misfortunes and is the most imperative element restricting sheep generation around the world (Prada et al., 2014; Valcarcel et al., 2015; Debebe et al., 2015). Anthelmintic are medications that demonstrate or systemically to kill grown-up helminths or formative stages that attack organs and tissues (Lou et al., 2014). *T. tubifex* is a provincially to oust worms from the gastro-intestinal tract cosmopolitan nauidid annelid sensu (Lou et al., 2014) representing one of the major components of the benthic fauna in freshwater communities (Schmelz et al., 2015). Also present in polluted waters, *T. tubifex* is widely used in laboratories for ecotoxicology research (Mendez et al., 2014) and as a model organism for the study of annelid development (Mendez et al., 2013). *T. tubifex* is characterized by considerable variability in its morphological features (Urbisz et al., 2015) and by a mixed reproductive strategy, with parthenogenesis (Marotta et al., 2014), self-fertilization (Gavrilov, 1935), and biparental reproduction through cross-mating (Sarker et al., 2015).

The aim of the present study is to identify the antidiarrheal activity by using different experimental methods and anthelmintic activity of methanol extract of *C. benghalensis* leaves using *T. tubifex* worm. However, no earlier studies have been conducted experimentally to characterize the antidiarrheal and anthelmintic effect of this leaves.

## MATERIALS AND METHODS

### Plant collection and identification

The leaves of *C. benghalensis* were collected from Chittagong, Bangladesh in the month of October, 2014. The leaves were taxonomically identified by Dr. Shaikh Bokhtear Uddin (Associate Professor, Department of Botany, University of Chittagong, Bangladesh). A voucher specimen (Accession No. Pharm P&D 71/09-15/30) was deposited at the Department of Pharmacy, International Islamic University Chittagong, Bangladesh for further reference.

### Extract preparation

The leaves were dried for a period of 2 weeks under shade and ground. The ground leaves (250 g) were soaked in sufficient amount of methanol for one week at room temperature with occasional shaking and stirring, and then filtered through a cotton plug followed by Whitman filter paper number 1. The solvent was evaporated under reduced pressure at room temperature to yield semisolid. The extract was then preserved in a refrigerator till

\*Corresponding author. E-mail: rmamun26@yahoo.com. Tel: +8801671322745.

further use.

### Experimental animals

Swiss albino mice, weighing about 25 to 30 g, were collected from Jahangir Nagar University, Savar, Bangladesh. The animals were provided with standard laboratory food and distilled water and maintained at natural day-night cycle having proper ventilation in the room. All the experiments were conducted in an isolated and noiseless condition. The study protocol was approved by the P&D Committee, Department of Pharmacy, International Islamic University of Chittagong, Bangladesh (Grant No. Pharm P&D 71/09-15/30). The animals were acclimatized to laboratory condition for 7 days prior to experimentation.

### Experimental worms

Experimental worms were collected from local aquarium shop. Then, authenticated by local zoologist (Authentication No. Wb2015).

### Chemicals and reagents

Loperamide (Square Pharmaceuticals Ltd., Bangladesh), castor oil (WELL's Health Care, Spain), normal saline solution (0.9% NaCl) and charcoal meal (10% activated charcoal in 5% gum acacia), and albendazole were used for anthelmintic activity tests. All other reagents were of analytical grade.

### Preparation of test doses

The extracts were suspended in the vehicle. Various strengths were prepared from a stock solution of 40 mg/ml. The solutions were prepared, and the freshly prepared solutions were administered orally.

### Acute toxicity study

For acute toxicity study, forty Swiss albino female mice were used. According to the method of Walum, mice were divided into four groups of five animals each (Walum, 1998). Different doses (1000, 2000, 3000 and 4000 mg/kg) of methanol extract of *C. benghalensis* leaves were administered by stomach tube. Then, the animals were observed for general toxicity signs.

### In vivo antidiarrheal activity

#### Castor oil-induced diarrhea

The experiment employed the method described by Awouters et al. (1978). Mice were fasted for 18 h before the test with free access to water and divided into four groups of five animals each. Group I treated as control (saline 2 ml/kg body weight intraperitoneally), Group II received standard drug (loperamide 5 mg/kg body weight p.o.), Groups III and IV received methanol extract of *C. benghalensis* (200 and 400 mg/kg body weight p.o.). Then, 1 h later, castor oil was administered orally to these animals to induce diarrhea. The mice's were then housed singly in cages lined with white blotting paper. The papers were changed every hour. The total number of both dry and wet feces excreted were counted every hour for a period of 4 h and compared with the control group. The total number of diarrheal feces of the control group was considered 100%.

### Castor oil induced enteropooling

Intraluminal fluid accumulation was determined by the method of Robert et al. (1976). Mice were fasted for 18 h and divided into five groups of four animals each. Group I served as control (saline 2 ml/kg body weight intraperitoneally), Group II received standard drug (loperamide 5 mg/kg body weight ip), Groups III and IV received methanol extract of *C. benghalensis* (200 and 400 mg/kg body weight po). Then, 1 h later, castor oil was administered orally to these animals to induce diarrhea. Two hours later, the mice were sacrificed by overdose of chloroform anesthesia, and the small intestine was ligated both at the pyloric sphincter and at the ileocecal junctions and dissected out. The small intestine was weighed. The intestinal contents were collected by milking into a graduated tube and the volume was measured. The intestines were reweighed and the differences between full and empty intestines were calculated.

### Gastrointestinal motility test

This experiment was carried out by the method described by Mascolo et al. (1994). Mice were fasted for 18 h and divided into four groups of five animals each. Castor oil was administered orally to these animals to induce diarrhea. One hour later, Group I received saline 2 ml/kg body weight intraperitoneally, Group II received standard drug (loperamide 5 mg/kg body weight ip), Groups III and IV received methanol extract of *C. benghalensis* (200 and 400 mg/kg body weight po). One hour after intraperitoneal administration of treatments, animals received 1 ml of charcoal meal (10% charcoal suspension in 5% gum acacia) orally. One hour later, the animals were sacrificed by overdose of chloroform anesthesia and the distance traveled by the charcoal meal from pylorus to caecum was measured and expressed as a percentage of the total distance of the intestine.

### In-vitro anthelmintic assay

The anthelmintic activity of methanol extract of *C. benghalensis* was carried out as per the procedure of Ajaiyeoba et al. (2001) with some minor modifications. The aquarium worm *T. tubifex* were used in the present study, because it has anatomical similarity and belongs to the same group of intestinal worm, that is, annelid (Verma et al., 2013; Raju et al., 2013; Rajagopal et al., 2013). The worms were collected from the local market of Chittagong, and average size of the worms, 2 to 2.5 cm in length were used for the study. The standard drug levamisole (1 mg/ml) and three different concentrations of methanol extract of *C. benghalensis* (2.5, 5 and 10 mg/ml) in double distilled water (Satish and Ravindra, 2009; Iqbal et al., 2001) were prepared freshly and used for the study of anthelmintic activity. One group composed of water and it was considered as the controlled group. The anthelmintic activity was determined at two different stages 'time of paralysis' and 'time of death' of the worms. Time for paralysis was noted when no movement of any sort could be observed except when the worms were shaken vigorously. Death was concluded when the worms lost their motility followed with fading away of their body colors (Grime et al., 2006). Death was also confirmed by dipping the worms in slightly warm water. The mortality of parasite was assumed to have occurred when all signs of movement had ceased (Temjenmongla and Yadav, 2005).

### Statistical analysis

The results are expressed as mean  $\pm$  standard error of the mean (SEM). Data were analyzed using one way factorial ANOVA tests



**Table 1.** Effect of methanol extract of leaves of *C. benghalensis* on castor oil (0.5 ml) induced diarrhea in Swiss albino mice.

Group	Treatment	Total number of feces	Inhibition of defecation (%)	Total number of diarrheal feces	Inhibition of diarrhea (%)
I	Saline (2 ml/kg p.o.)	13.4±0.245	-	5.8±0.2	-
II	Loperamide (5 mg/kg p.o.)	5.4±0.4 <sup>c</sup>	59.70±2.99	2.2±0.374 <sup>b</sup>	62.07±6.45
III	MECB (200 mg/ml p.o.)	8.8±0.2 <sup>c</sup>	34.33±1.49	4.2±0.2 <sup>b</sup>	27.59±3.45
IV	MECB (400 mg/ml p.o.)	7±0.547 <sup>c</sup>	47.76±4.09	3.2±0.2 <sup>c</sup>	55.17±3.45

Values are mean ± SEM ( $n = 5$ ); <sup>a</sup> $p < 0.05$ , <sup>b</sup> $p < 0.01$  and <sup>c</sup> $p < 0.001$ , Dunnett test as compared to negative control (saline). Statistical representation of the total number of feces and total number of diarrheal feces by *C. benghalensis* methanol extract, positive antidiarrheal control (Loperamide, 5 mg/kg p.o.) processed by paired t-test analysis (Dennett's test). Data were processed by paired t-test analysis by using SPSS for windows, version 16.0.

**Table 2.** Effect of methanolic extract of leaves of *C. benghalensis* on castor oil induced enteropooling in Swiss albino mice.

Group	Treatment	Volume of intestinal content (ml)	Weight of intestinal content (g)	Inhibition of intestinal content (%)
I	Saline (2 ml/kg p.o.)	0.24±0.013	0.628±0.033	-
II	Loperamide (5 mg/kg p.o.)	0.13±0.001	0.316±0.002 <sup>b</sup>	48.2±0.40
III	MECB (200 mg/ml p.o.)	0.17±0.007	0.408±0.02 <sup>b</sup>	33.11±3.64
IV	MECB (400 mg/ml p.o.)	0.15±0.005	0.334±0.01 <sup>b</sup>	45.25±1.84

Values are mean ± SEM ( $n = 5$ ); <sup>a</sup> $p < 0.05$ , <sup>b</sup> $p < 0.01$  and <sup>c</sup> $p < 0.001$ , Dunnett test as compared to negative control (saline). Statistical representation of the weight of intestinal content (g) by *C. benghalensis* methanol extract, positive antidiarrheal control (Loperamide, 5 mg/kg p.o.) processed by paired t-test analysis (Dennett's test). Data were processed by paired t-test analysis by using SPSS for windows, version 16.0.

using SPSS Data Editor for Windows, Version 16.0 (SPSS Inc., USA) followed by Dennett's tests on each group except the control group for anthelmintic. The results obtained were compared with the negative control group for antidiarrheal activity and  $P < 0.05$ ,  $P < 0.01$  and  $P < 0.001$  was considered to be statistically significant in Dennett's tests. Statistical program GRAPHPAD PRISM® (version 6.00; GraphPad Software Inc., San Diego, CA, USA) was used for graphical presentation.

## RESULTS

### Acute toxicity test

None of the animals showed behavioral, neurological or physical changes characterized by symptoms, such as reduced motor activity, restlessness, convulsions, coma, diarrhea and lacrimation at the limit dose of 4000 mg/kg for methanol extract of *C. benghalensis* during the observation period. In addition, no mortality was observed at the test dose. Thus, the median lethal dose ( $LD_{50}$ ) of the plant extract was found to be greater than 4000 mg/kg.

### In vivo antidiarrheal activity

#### Castor oil-induced diarrhea

In the castor oil-induced diarrhea conduct experiment, the leaves extract of *C. benghalensis* produced a noticeable

antidiarrheal result in the mice's, as shown in Table 1. At doses of 200 and 400 mg/kg, the extract produced significant ( $p < 0.01$ ) defecation. The total number of wet feces produced upon administration of castor oil decreased ( $4.2 \pm 0.2$ , at 200 mg/kg and  $3.2 \pm 0.2$ , at 400 mg/kg) compared to the control group ( $5.8 \pm 0.2$ ) while loperamide decreased to  $2.2 \pm 0.374$  at the dose of 5 mg/kg.

#### Castor oil induced enteropooling

Castor oil caused accumulation of water and electrolytes in intestinal loop. Treatment with the *C. benghalensis* extract (200 and 400 mg/kg) produced a significant and dose-dependent reduction in intestinal weight and volume (Table 2). The intestinal volume was decreased by  $33.11 \pm 3.64$  and  $45.25 \pm 1.84\%$  at doses 200 and 400 mg/kg, respectively. The standard drug, loperamide (5 mg/kg), also significantly inhibited ( $p < 0.01$ ) intestinal fluid accumulation ( $48.2 \pm 0.40\%$ ).

#### Gastrointestinal motility test

The consequence of *C. benghalensis* extract on the intestinal transit is shown in Table 3. All doses of the extracts successful produced significant alteration in the percent of intestinal motility compared to the negative control. The negative control (saline) resulted in  $84.85 \pm 2.88\%$

**Table 3.** Effect of methanolic extract of leaves of *C. benghalensis* on charcoal induced gut transit changes in Swiss albino mice.

Group	Treatment	Intestine cross by marker (%)	Inhibition (%)
I	Saline (2 ml/kg p.o.)	84.85±2.88	
II	Loperamide (5 mg/kg po)	45.35±1.72 <sup>c</sup>	43.6±2.14
III	MEHO (200 mg/ml po)	57.95±0.71 <sup>b</sup>	27.92±0.88
IV	MEHO (400 mg/ml po)	51.99±1.2 <sup>c</sup>	35.34±1.5

Values are mean ± SEM (n = 5); <sup>a</sup>p < 0.05, <sup>b</sup>p < 0.01 and <sup>c</sup>p < 0.001, Dunnett test as compared to negative control (saline). Statistical representation of the % of intestine cross by marker by *C. benghalensis* methanol extract, positive antidiarrheal control (Loperamide, 5 mg/kg p.o.) processed by paired t-test analysis (Dennett's test). Data were processed by paired t-test analysis using SPSS for windows, version 16.0.

**Table 4.** Anthelmintic activity of methanol extract of *C. benghalensis*.

Treatment	Time taken for paralysis (min)	Time taken for Death (min)
Control (Water)	0	0.00
Levamisole (1 mg/ml)	3.3±0.38	6.5±0.76
<i>C. benghalensis</i> (10 mg/ml)	12.38±0.43 <sup>b</sup>	21.39±0.50 <sup>b</sup>
<i>C. benghalensis</i> (5 mg/ml)	16.25±0.48 <sup>b</sup>	45.64±1.01 <sup>b</sup>
<i>C. benghalensis</i> (2.5 mg/ml)	22.17±0.49 <sup>b</sup>	57.25±1.18 <sup>b</sup>

Values are mean ± SEM, (n = 3); <sup>a</sup>p < 0.05, <sup>b</sup>p < 0.01 and <sup>c</sup>p < 0.001, Dennett's test as compared to positive control (Levamisole, 1 mg/ml). Statistical representation of the effective paralysis and dead time by *C. benghalensis* methanol extract, positive anthelmintic control (Levamisole, 1 mg/ml) processed by paired t-test analysis (Dennett's test). Data were processed by paired t-test analysis using SPSS for windows, version 16.0.

intestinal motility by the marker-charcoal meal. The 200 and 400 mg/kg oral dose of the extracts of *C. benghalensis* exhibited 57.95±0.7 and 51.99±1.2<sup>c</sup> intestinal motility (Table 3). And the extracts significantly inhibited 27.92±0.88 and 35.34±1.5 % at all doses in intestinal motility. However, the standard drug, loperamide (5 mg/kg) demonstrated a significant inhibition (43.6±2.14%) in intestinal motility.

### Anthelmintic activity

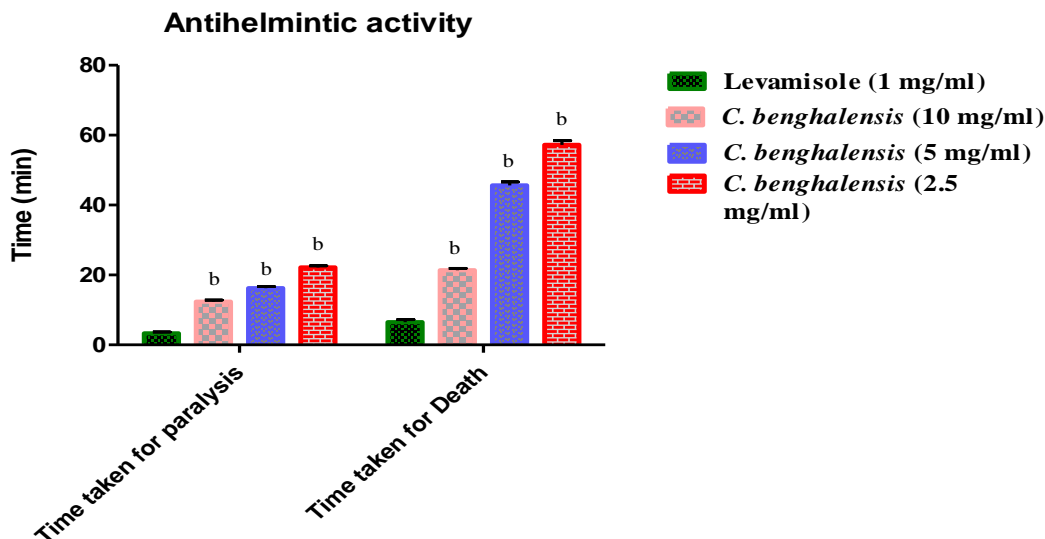
Consequences of the study were recorded as shown in Table 4 and Figure 1 as in the form of time required to get the following attacks of paralysis and at the end time required for complete death of parasite. From the aforementioned study, it was seen that the methanolic extract showed dose dependent anthelmintic activity as compared to a standard drug levamisole. Methanol extract *C. benghalensis* show the time taken for paralysis (12.38, 16.25, and 22.17 min) and time taken for death (21.39, 45.64, and 57.25 min) at dose 10, 5, and 2.5, respectively compared to a standard drug levamisole (1 mg/ml) that showed the time taken for paralysis (3.3±0.38 min) and time taken for death (6.5±0.76 min).

### DISCUSSION

It is generally realized that castor oil is metabolized into

ricinoleic corrosive in the gut which is in charge of looseness of the bowels generation (Darracq et al., 2015). The peristaltic action of small digestive tract is expanded if ricinoleate is introduced in the small digestive system, thus Na<sup>+</sup> and Cl<sup>-</sup> porousness changed in intestinal mucosa (Tariq et al., 2015). Emission of endogenous prostaglandin is empowered additionally by ricinoleate (Beubler and Juan, 1979; Sadraei et al., 2014). The leaves extract of *C. benghalensis* at 200 and 400 mg/kg measurements showed enormous decrease of the quantity of diarrheal and total feces which may be because of the initiation of prostaglandin biosynthesis with resultant abatement in emission of liquid into the lumen or may be because of advancement and ingestion of water and electrolytes in the gut. The standard drug, loperamide (5 mg/kg) also produced statistically significant (p<0.01) diarrheal inhibition (62.07%). The intestinal volume was decreased by 33.11 and 45.25% at doses of 200 and 400 mg/kg, respectively. The aforementioned hypothesis was further upheld by the inhibitory activity of the extract on intestinal charcoal supper motility. Methanol extract of *C. benghalensis* smothered the propulsive development or travel of charcoal supper through the gastrointestinal tract which fundamentally shows that the extract may have the probability to decrease the recurrence of stooling in diarrheal conditions.

Anthelmintic medications act quickly and specifically on neuromuscular transmission of nematodes. Three major



**Figure 1.** Anthelmintic activity of methanol extract of *C. benghalensis*. Values are mean  $\pm$  SEM ( $n = 3$ ); <sup>a</sup> $p < 0.05$ , <sup>b</sup> $p < 0.01$  and <sup>c</sup> $p < 0.001$ , Dennett's test as compared to positive control (Levamisole, 1 mg/ml). Statistical representation of the effective paralysis and dead time by *C. benghalensis* methanol extract, positive anthelmintic control (Levamisole, 1 mg/ml) processed by paired t-test analysis (Dennett's test). Data were processed by paired t-test analysis using SPSS for windows, version 16.0.

groups of helminths (worms), the nematodes, trematodes, and cestodes- infect human (Mwale and Masika 2015; Pawluk et al., 2015; Romero et al., 2014). Nematodes are elongated roundworms that possess a complete digestive system, including both mouth and anus. These cause the infections of the intestine as well as the blood and tissues. The trematodes (flukes) are leaf-shaped flatworms that are generally characterized by the tissues they infect. For example, they may be categorized as liver, lung, intestinal, or blood flukes. The cestodes or true tapeworms typically have a flat, segmented body and are attach to the hosts intestine. Levamisole are agonists at nicotinic acetylcholine receptors of nematode muscle and reason spastic loss of motion.

The methanol extract of *C. benghalensis* showed anthelmintic activity at a concentration of 10, 5 and 2.5 mg/ml. The anthelmintic effect of the extract is comparable with that of the effect produced by the standard drug levamisole (1 mg/ml) which also produced statistically significant ( $p < 0.01$ ) anthelmintic activity time taken for paralysis (3.3 min) and time taken for death (6.5 min). The activity was concentration dependent. The plant possesses significant anthelmintic activity at 10, 5, and 2.5 mg/ml concentration measured by time taken for paralysis (12.38, 16.25, and 22.17 min) and death of the earth worm (21.39, 45.64, and 57.25 min) in that order.

## Conclusion

This study underpins that the methanol extract of *C. benghalensis* leaves are the planned sources of

antidiarrheal and anthelmintic specialists in the conventional drug framework. In this case, studies are obliged to assist to distinguish the dynamic constituent(s) of the portions to comprehend the pharmacological activity of the antidiarrheal and anthelmintic impacts. The discoveries propose the legitimacy of the much lauded impact of *C. benghalensis* as antidiarrheal and anthelmintic specialists in expected home grown medication.

## Conflict of Interests

The authors have not declared any conflict of interests.

## ACKNOWLEDGEMENT

The authors express their thanks to the Jahangir Nagar University, Savar, Dhaka, Bangladesh, for providing experimental mice.

## REFERENCES

- Adjanohoun E (1993). Contribution to ethnobotanical and foristic studies in Uganda. OUA/CSTR, Lagos. From the data bank PHARMEL 2 (ref. HP 10).
- Ajaiyeoba EO, Onocha PA, Olarenwaju OT (2001). *In vitro* anthelmintic properties of *Buchholzia coriacea* and *Gynandropsis gynandra* extract. Pharm. Biol. J. 39:217-220.
- Awouters F, Niemegeers CJE, Lenaerts FM, Janseen PAJ (1998). Delay of castor oil diarrhea in rats; a new way to evaluate inhibitors of prostaglandin biosynthesis. J. Pharmacol. 30:41-45.
- Beubler E, Juan H (1979). Effect of ricinoleic acid and other laxatives on net water flux and prostaglandin E release by the rat colon. J. Pharm. Pharmacol. 31:681-685.

- Chowdhury F, Khan IA, Patel S, Siddiq AU, Saha NC, Khan AI (2015). Diarrheal illness and healthcare seeking behavior among a population at high risk for diarrhea in Dhaka, Bangladesh. *PLoS One* 10(6):e0130105.
- Chowdhury TA, Hasanat A, Jakaria M, Mostafa ATMK, Kabir MSH, Hossain S, Mamur A, Hossain MM (2015). Thrombolytic and cytotoxic activity of methanolic extract of *Commelina benghalensis* (Family: Commelinaceae) Leaves. *J. Sci. Innovates Res.* 4(2):100-104
- Darracq MA, Cullen J, Rentmeester L, Cantrell FL, Ly BT (2015). Orbeez: The magic water absorbing bead-risk of pediatric bowel obstruction. *Pediatric emergency care* 31(6):416-418.
- Debebe Y, Tefera M, Mekonnen W, Abebe D, Woldekidan S, Abebe A (2015). Evaluation of anthelmintic potential of the Ethiopian medicinal plant *Embelia schimperi* Vatke *in vivo* and *in vitro* against some intestinal parasites. *BMC Complementary Altern. Med.* 15:187-015-0711-7.
- Dooley LA, Froese EA, Chung YT, Burkman EJ, Moorhead AR, Ardelli BF (2015). Host ABC transporter proteins may influence the efficacy of ivermectin and possibly have broader implications for the development of resistance in parasitic nematodes. *Exp. Parasitol.* 4(3):157.
- Effets spasmodiques des extraits aqueux et au mélange méthanol/chlorure de méthylène (1:1) des feuilles de *Gmelina arborea* (Verbenaceae) sur la motricité intestinale de rat. *Cameroon J. Exp. Biol.* 2:31-38.
- Galheigo MR, Prado LC, Mundin AM, Gomes DO, Chang R, Lima AM (2015). Antidiarrhoeal effect of *Eugenia dysenteric* DC (myrtaceae) leaf essential oil. *Natural Product Res.* 2:1-4.
- Gavrilov C (1935). Contributions a l'étude de l'autofécondation chez les oligochètes. *Acta Zoo.* 16:21-64.
- Grime AS, Bhalke RD, Ghogare PB, Tambe VD, Jadhav RS, Nirmal SA (2006). Comparative *in vitro* anthelmintic activity of *Mentha piperita* and *Lantana camara* from Western India. *Dhaka University J. Pharm. Sci.* 5(1-2):5-7.
- Guerrant RL, Van Gilder T, Steiner TS, Theilman MN, Slutsker L (2001). Practice guidelines for the management of infectious diarrhea. *J. Infect Dis.* 32:331-351.
- Hasan SMR, Hossain MM, Akter R, Jamila M, Mazumder MEH, Rahman S (2009). Sedative and anxiolytic effects of different fractions of the *Commelina benghalensis* Linn. *Drug discoveries ther.* 3:221-227.
- Hasan SMR, Hossain MM, Faruque A, Mazumder MEH, Rana MS, Akter R, Alam MA (2008). Comparison of antioxidant potential of different fractions of *Commelina benghalensis* Linn. *J. Life Sci.* 20:9-16.
- Hong D, DePhillips RA (2000). *Commelina diffusa*, in *Flora of China*, Wu ZY, Raven PH, Hong DY (eds.), Beijing, Science press, St. Louis, Missouri botanical garden press. p36.
- infections on small ruminant farms. *Vet. Parasitol.* 211(1-2):71-79.
- Iqbal Z, Nadeem QK, Khan MN, Akhtar MS, Waraich FN (2001). *In vitro* Anthelmintic Activity of *Allium sativum*, *Zingiber officinale*, *Cucurbita mexicana* and *Ficus religiosa*. *Int. J. Agric. Biol.* 3(4):454-457.
- Jayvir A, Minoo P, Gauri B, Ripal K (2007). *Natural Heals: A glossary of selected indigenous medicinal plant of India*, 2<sup>nd</sup> ed, SRIST Innovations, Ahmadabad, India. p22.
- Kirtikar KR, Basu BD (1980). *Industrial Medicinal Plants*, 2nd ed. 2532-2541.
- Lou JQ, Yang DY, Cao YQ, Sun PD, Zheng P (2014). Physiological responses of tubificidae to heavy metal chromium stress. *Bian Ji* 35(11):4205-4211.
- Manandhar N, Sanjay P (2000). *Plants and people of Nepal*, Timber Press, Nepal.
- Marotta R, Crottini A, Raimondi E, Fondello C, Ferraguti M (2014). Alike but different: The evolution of the tubifex tubifex species complex (annelida, clitellata) through polyploidization. *BMC Evol. Biol.* 14(1):73.
- Mascolo N, Izzo AA, Autore G, Barbato F, Capasso F (1994). Nitric oxide and castor oil- induced diarrhea. *J. Pharmacol. Exp. Ther.* 268:291-295.
- Mbazima VG, Mokgotho MP, February F, Rees DJG, Mampuru LJ (2008). Alteration of Bax-to-Bcl-2 ratio modulates the anticancer activity of methanolic extract of *Commelina benghalensis* (Commelinaceae) in Jurkat T cells. *Afr. J. Biotechnol.* 7:3569-3576.
- Mendez FL, De JM, Bervoets L (2014). Influences of sediment geochemistry on metal accumulation rates and toxicity in the aquatic oligochaete *Tubifex tubifex*. *Aquatic Toxicol.* 157:109-119.
- Mendez FL, Martinez MM, Rodriguez P (2013). Toxicity and critical body residues of cd, cu and cr in the aquatic oligochaete tubifex tubifex (muller) based on lethal and sublethal effects. *Ecotoxicology* 22(10):1445-1460.
- Mwale M, Masika PJ (2015). *In vivo* anthelmintic efficacy of aloe ferox, agave sisalana, and gunnera perpensa in village chickens naturally infected with heterakis gallinarum. *Trop. Anim. Health Prod.* 47(1):131-138.
- Okello J, Ssegawa P (2007). Medicinal plants used by communities of Ngai Subcounty, Apac District, northern Uganda. *Afr. J. Ecol.* 45:76-83.
- Parehs J, Chanda SV (2008). Antibacterial activity of aqueous and alcoholic extracts of 34 Indian medicinal plants against some *Staphylococcus* species. *Turk. J. Biol.* 32:63-71.
- Patzi VS, Zaidi MB, Perez MI, Leon CM, Michel AA, Chaussabel D (2015). Diarrheagenic escherichia coli carrying supplementary virulence genes are an important cause of moderate to severe diarrhoeal disease in Mexico. *PLoS Neglected Tropical Dis.* 9(3):e0003510.
- Pawluk SA, Roels CA, Wilby KJ, Ensom MH (2015). A review of pharmacokinetic drug-drug interactions with the anthelmintic medications albendazole and mebendazole. *Clin. Pharm.* 54(4):371-383.
- Prada JCJ, Stear MJ, Mair C, Singleton D, Stefan T, Stear A (2014). An explicit immunogenetic model of gastrointestinal nematode infection in sheep. *J. Royal Society* 11(99):109.
- Qaiser M, Jafri SMH (1975). *Commelina benghalensis*, in *Flora of Pakistan*, Ali SI, Qaiser M (eds.), St. Louis: University of Karachi and Missouri botanical garden, p10.
- Rahman GMS, Haque N, Rashid A (1999). Cytotoxic activity of *Commelina benghalensis* Linn., using brine shrimp lethality bioassay. *Bangladesh J. Physiol. Pharmacol.* 15:62-65
- Rajagopal PL, Kiron SS, Sreejith KR, Premaletha K (2013). Anthelmintic studies on the whole plant of *Biophytum sensitivum* (L). *Int. J. drug formulation Res.* 4(5): 45-47.
- Raju GS, Moghal MR, Dewan SM, Amin MN, Billah M (2013). Characterization of phytoconstituents and evaluation of total phenolic content, anthelmintic, and antimicrobial activities of solanum violaceum Ortega. *Avicenna J. Phytomed.* 3(4):313-320.
- Raju M, Varakumar S, Lakshminarayana R, Krishnakantha TP, Baskaran V (2007). Carotenoid composition and vitamin A activity of medicinally important green leafy vegetables. *Food Chem.* 101:1598-1605.
- Robert A, Nezamis JE, Lancaster C, Hanchar AJ, Klepper MS (1976). Enteropooling assay: A test for diarrhea produced by prostaglandins. *Prostaglandins* 11:809-828.
- Romero MC, Navarro MC, Martin-Sanchez J, Valero A (2014). Peppermint (*Mentha piperita*) and albendazole against anisakiasis in an animal model. *Tropical Med. Int. Health* 19(12):1430-1436.
- Sadraei H, Ghanadian M, Asghari G, Madadi E, Azali N (2014). Antispasmodic and antidiarrheal activities of 6-(4-hydroxy-3-methoxyphenyl)-hexanoic acid from *Pycnocyca spinosa* Decne. *exBoiss. Res. Pharm. Sci.* 9(4):279-286.
- Sarker S, Kallert DM, Hedrick RP, El-Matbouli M (2015). Whirling disease revisited: Pathogenesis, parasite biology and disease intervention. *Diseases of aquatic organisms* 114(2):155-175.
- Satish BK, Ravindra A (2009). *Fursule* investigation of *in vitro* anthelmintic activity of *Thespesia lampas*. *Asian J. Pharm. Clin. Res.* 2:69-70.
- Schmelz RM, Jocque M, Collado R (2015). Microdrile oligochaeta in bromeliad pools of a Honduran cloud forest. *Zoo* 3947(4):508-526.
- Sharma MC, Sharma S (2010). Preliminary phytochemical and antimicrobial investigations of the aqueous extract of *Ixora coccinea* Linn and *Commelina benghalensis* L. on Gram-positive and Gram-negative microorganisms. *Middle-East J. Sci. Res.* 6:436-439.
- Ssenyonga M, Brehony E (1993). Herbal medicine-its use in treating

- some symptoms of AIDS. Int. Conf. AIDS, 9: 75-75
- Tabuti JR, Lye KA, Dhillion SS (2003). Traditional herbal drugs of Bulamogi, Uganda: plants, use and administration. J. Ethnopharmacol. 88:19-44.
- Tariq A, Mussarat S, Adnan M, Abd EF, Hashem A, Alqarawi AA (2015). Ethnomedicinal evaluation of medicinal plants used against gastrointestinal complaints. BioMed. Res. Int. 892947.
- Temjenmongla, Yadav AK (2005). Anticestodal efficacy of folklore medicinal plants of Naga tribes in Northeast India. Afr. J. Tradit. 2(2):129-133.
- Urbisz AZ, Chajec L, Swiatek P (2015). The ovary of tubifex tubifex (clitellata, nauididae, tubificinae) is composed of one, huge germ-line cyst that is enriched with cytoskeletal components. PloS One. 10(5):e0126173.
- Valcarcel F, Aguilar A, Sanchez M (2015). Field evaluation of targeted selective treatments to control subclinical gastrointestinal nematode infections on small ruminant farms." Vet. Parasitol. (2015).
- Van SF, Nkwanyana MN, De WH (2015). Antimicrobial evaluation of plants used for the treatment of diarrhea in a rural community in northern maputaland, KwaZulu-Natal, South Africa. BMC Complement. Alternat. Med. 15:53-015-0570-2.
- Verma VK, Sarwa K, Kumar A (2013). Anthelmintic Activity of Fruit Peel and Root Extracts of *Trapa natans*. Acad. J. Plant Sci. 6(2):73-76.
- Walum E (1998). Acute oral toxicity. Environ. Health perspectives 106(2):497-503.
- Wansi SL, Nguenefack TB, Watcho P, Ndam A, Kamanyi A (2007). Effects plasmogéniques des extraits aqueux et au mélange méthanol/chlorure de méthylène (1: 1) des feuilles de *Gmelina arborea* (Verbenacée) sur la motricité intestinale de rat. Cameroon J. Exp. Biol. 2(1).
- Yusuf M, Wahab MA, Chowdhury JW, Japripa BB (1994). Medical Plants of Bangladesh, BCSIR, Chittagong Laboratory, Bangladesh. P 73.
- Zavala MA, Perez S, Perez C, Vargal R, Perez RM (1998). Antidiarrhoeal activity of *Waltheria americana*, *Commelins cuelestris* and *Alternanthera repens*. J. Ethnopharmacol. 61:41-47.
- Zheng FH, Wei P, Huo HL, Xing XF, Chen FL, Tan XM (2015). Neuroprotective effect of gui zhi (ramulus cinnamomi) on ma huang-(herb ephedra-) induced toxicity in rats treated with a ma huang-gui zhi herb pair. Evid. Based Complement. Alternat. Med. 9(13):461.

*Full Length Research Paper*

## Potential impact of urocortin I on sperm count, motility and sex hormone profiles in normal adult rats

Hany A. El kattawy<sup>1</sup>, Mahmoud A. Naga<sup>2,3</sup>, Ahmed Shata<sup>3\*</sup> and Mohamed A. Elbaky<sup>4</sup>

<sup>1</sup>Department of Physiology, Faculty of Medicine, Zagazig University, Zagazig, Egypt.

<sup>2</sup>Department of Clinical Pharmacology and Toxicology, AL-Qunfudh Medical college, Umm Al-Qura university, Makka, Saudia Arabia.

<sup>3</sup>Department of Clinical Pharmacology, Mansoura University, Mansoura, Egypt.

<sup>4</sup>Department of Biochemistry, Faculty of Medicine, Minia University, Egypt.

Received 22 July 2016; Accepted 19 August 2016

**Urocortin I (Ucn I), Ucn II and Ucn III are corticotropin-releasing factor (CRF) - like peptides. Ucn I has a high affinity for CRF<sub>1</sub> and CRF<sub>2</sub> receptors. The function of Ucn in male reproduction has not yet been elucidated. Ucn I is present in rat testis with lower levels of Ucn II and III mRNA gene expressions. Therefore, it is suggested that Ucn I may play a role in the regulation of male reproductive action. This study is designed to find out the endocrine and reproductive effects of Ucn I in normal adult male rats through investigation of exogenous Ucn I effects on epididymal sperm count, sperm motility, testicular weight and sex hormone profiles. 24 normal adult male albino rats of 175 to 200 gm initial body weight were implemented in this study. Randomly, the rats were subdivided into four equal groups. Group (I): Vehicle-treated group in which control rats received 0.1 mL of normal saline for 42 days as Ucn I-treated groups. Rats in group II were divided into three equal subgroups. Group (IIA), (IIB) and (IIC): Ucn I-treated groups in which rats were given daily intraperitoneal injections of rat Ucn I at doses of 5, 10 and 20 µg/kg body weight respectively for 42 days. Rats were weighed. Serum testosterone and levels of Follicle-stimulating hormone (FSH) and Luteinizing hormone (LH) were measured, in addition to epididymal sperm count, sperm motility and testicular weight. Administration of Ucn I in dose dependant manner induced significant reducing effects upon body weight, testes weight, epididymal sperm count, sperm motility and serum testosterone levels of normal adult rats. Ucn I may possibly affect male infertility by induction of a significant decrease in epididymal sperm count, motility and serum testosterone levels that might be due to a direct effect of Ucn I on testicular tissue and the germ cells without direct involvement of hypothalamic-pituitary-gonadal axis.**

**Key words:** Urocortin I, epididymal sperm count, sperm motility, testosterone, male rat.

### INTRODUCTION

Urocortin (Ucn) I, II and III and corticotropin releasing hormone (CRH) are peptide hormones that belong to the

\*Corresponding author. E-mail: ahmedmhes@mans.edu.eg.

corticotropin releasing hormone family of peptides. Ucn I has a high affinity for CRF<sub>1</sub> and CRF<sub>2</sub> receptors. However, Ucn II and Ucn III show selective CRF<sub>2</sub> affinity (Hsu and Hsueh, 2001, Karteris et al., 2004).

Ucn I, II and III have been detected in the central nervous system (Bittencourt et al., 1999) as well as in many peripheral tissues (Kageyama et al., 1999; Lewis et al., 2001, Oki and Sasano, 2004). Ucn's have roles in diverse physiologic processes such as regulation of the cardiovascular, gastrointestinal, reproductive, immune functions, body weight, food intake, and behavioral responses to stressors (Rodriguez et al., 1983, Fekete and Zorrilla, 2007).

Ucn I is composed of 40 amino acids with 45% sequence identity to CRF and 63% sequence identity to fish urotensin. It was first isolated from rat midbrain (Vaughan et al., 1995; Zhao et al., 1998). The major site of brain Ucn I synthesis is the Edinger-Westphal nucleus (Bittencourt et al., 1999).

The reproductive function of Ucn's in male is still unclear. However, both Ucn mRNA and peptide were expressed in mature spermatozoa. Moreover, the expressions of CRF<sub>1</sub> and CRF<sub>2</sub> receptors in spermatocytes and spermatogonia respectively suggested that Ucn's may play a role in the germ cell division, differentiation and spermatogenesis (Tao et al., 2007; Rivier, 2008).

Ucn I gene is located in rat testis with lower detectable levels of Ucn II and III mRNA gene expressions indicating the predominance of Ucn I signals. Ucn I gene expression appeared restricted to interstitial Leydig cells (Lee et al., 2011). Hence, the testis is a main target for Ucn I interactions. The prostate is a main source of local Ucn I. The secreted Ucn in the seminal fluid could activate the myometrial contractility in the female genital tract. So, it is suggested that Ucn I may participate in the physiology of fertilization, pregnancy and parturition (Petraglia et al., 1999; Yutaka and Hironobu., 2004, Lee et al., 2011).

Therefore, this study was designed to find out the endocrinal and reproductive function of Ucn I in male through evaluating the effects of exogenous Ucn I on epididymal sperm count, motility and sex hormone profiles in normal adult rats.

## MATERIALS AND METHODS

24 adult albino male rats weighing 175 - 200 gm were obtained from the animal house of Faculty of Veterinary Medicine- Zagazig University. They were put under well controlled light and temperature conditions before experiments in the physiology animal house in Faculty of Medicine - Zagazig University. Animals were not manipulated except for feeding or cleaning of houses and were allowed to eat and drink ad libitum. Animals were handled with principles for the care and use of research as adopted by the National Institutes of Health and the approval from Animal Ethic Committee of the institution (Egypt). After one week of acclimatization, the rats were randomly and equally divided into four

groups. Group (I): Vehicle-treated group in which control rats received 0.1 mL of normal saline for 42 days. Rats in group II were divided into three equal subgroups. Group (IIA), (IIB) and (IIC): Ucn I-treated groups in which rats were given daily intraperitoneal injections of rat Ucn I at doses of 5, 10 and 20 µg / kg body weight respectively for 42 days (Haron et al., 2010). Rat Ucn I (1 mg, powder form, Sigma Chemical Co., St. Louis, USA) was dissolved with 1% acetate solution and frozen at -80°C until use (Kihara et al., 2001). All injections were performed intraperitoneally in a volume of 0.1 ml/rat.

## Samples collection

24 h after the last injection of Ucn I, blood samples were taken from retro-orbital venous plexus. Serum was separated by centrifugation of blood at 3000 rpm for 20 minutes and kept deep frozen at (-20°C). Serum Follicle-stimulating hormone (FSH) and Luteinizing hormone (LH) and testosterone levels were measured. Under mild ether anesthesia, the animals were sacrificed by cervical dislocation. Testes were weighed and epididymis was used for the evaluation of sperm parameters. Repeated freezing and thawing of samples were avoided.

## Hormonal assays

Serum FSH levels were measured using FSH enzyme immunoassay test kits (BioCheck, CA 94404) according to the method described by Rebar et al. (1982). Both serum LH levels and testosterone levels were measured using LH enzyme immunoassay test kits and testosterone enzyme immunoassay test kits respectively (BioCheck, CA 94404) (Tietz, 1995). All are measured by spectrophotometer (spectronic 3000 Array, Germany) at 450 nm.

## Sperm count and motility analysis

For each rat, the right epididymis was dissected, removed and minced in 2 ml of Hank's buffer salt solution (HBSS) (Sigma-Aldrich Co.-USA) at 37°C (Idris et al., 2012). The cauda epididymis sperm was determined using the standard hemocytometer method after 5 min incubation at 37°C, then the epididymal fluid was drawn up to the 0.5 mark of pipette White Blood Cell (WBC) and the semen diluting fluid (sodium bicarbonate 5 g, formalin 1 ml, distilled water 99.0 ml) was drawn up to '11' mark, and subsequently mixed well. One drop was kept in haemocytometer chamber in humid place for 1 h. Then, under the light microscope, the number of spermatozoa in the appropriate haemocytometer squares was counted according to the formula; sperm count = No. of spermatozoa counted x dilution factor x volume factor/ No. of areas counted (Belsey et al., 1980), the sperm concentration refers to the number of spermatozoa / ml fluid was gauged.

Using the number of live sperm cells over the total number of sperm cells (either motile which displayed some movement or non-motile that not move at all), the sperm motility percentage was calculated (Khaki et al., 2009).

## Statistical analysis

The statistical analysis of results was done by using Statistical Package for Social Science (SPSS) program, version 18, for windows XP professional. The biochemical data were expressed as Mean ± SD. Statistical analysis were performed using one way analysis of variance (ANOVA) followed by post-hoc multiple comparisons (Tukey test). P value < 0.05 was considered

**Table 1.** Serum LH, FSH, T levels in Ucn I-treated rats and controls after a duration of 42 days.

Variable	FSH ( $\mu\text{UL/mL}$ )	LH ( $\mu\text{UL/mL}$ )	T (ng/mL)
Group I (Control rats)	0.43 $\pm$ 0.04	0.41 $\pm$ 0.07	4.39 $\pm$ 0.79
Group IIA	0.40 $\pm$ 0.03	0.39 $\pm$ 0.05	3.5 $\pm$ 0.47
Group IIB	0.38 $\pm$ 0.04	0.40 $\pm$ 0.04	2.7 $\pm$ 0.34 <sup>p1,p2</sup>
Group IIC	0.37 $\pm$ 0.04	0.37 $\pm$ 0.073	1.87 $\pm$ 0.41 <sup>p1, p2</sup>

p1 = significant difference when compared with negative control group, p2 significant when compared with group IIA, p3 when compared with group IIB

**Table 2.** Epididymal sperm count, motility and testis weight in Ucn I-treated rats and controls after a duration of 42 days.

Variable	Epididymal sperm count (millions/ml)	Total sperm motility (%)	Progressive motility (%)	Testis weight (g)
Group I (Control rats)	53.1 $\pm$ 5.9	85.9 $\pm$ 9.8	78.9 $\pm$ 8.86	1.84 $\pm$ 0.3
Group IIA	43.7 $\pm$ 5.5 <sup>p1</sup>	70.3 $\pm$ 7.4 <sup>p1</sup>	61.72 $\pm$ 8.1 <sup>p1</sup>	1.48 $\pm$ 0.17 <sup>p1</sup>
Group IIB	40.7 $\pm$ 5.5 <sup>p1</sup>	67.1 $\pm$ 10.6 <sup>p1</sup>	59.4 $\pm$ 11.1 <sup>p1</sup>	1.17 $\pm$ 0.16 <sup>p1</sup>
Group IIC	35.2 $\pm$ 6.1 <sup>p1</sup>	60.1 $\pm$ 9.2 <sup>p1</sup>	56.31 $\pm$ 11.35 <sup>p1</sup>	0.87 $\pm$ 0.13 <sup>p1</sup>

p1 = significant difference when compared with negative control group, p2 significant when compared with group IIA, p3 when compared with group IIB

Total Motility = Progressive Motility + Non-progressive Motility; Progressive motility: sperms moving actively, either linearly or in large circle regardless of speed; Non-progressive motility: all other motility patterns with an absence of progression (i.e. swimming in a small circle or when only a flagellar beat can be observed); Immotile sperms: no movement.

statistically significant at confidence interval 95 %.

## RESULTS

### Hormone assays

Table 1 shows the effects of Ucn I on hormones in the studied groups. Following treatment with different doses of Ucn I, the results showed no significant differences were evident in serum FSH and LH levels between rats given Ucn I and their age-matched controls ( $p1 > 0.05$ ). However, T was significantly lower in rats treated with 10 and 20  $\mu\text{g} / \text{kg}$  ( $p2 < 0.04$ ,  $p2 < 0.001$  respectively) compared to those treated by 5  $\mu\text{g} / \text{kg}$  of Ucn I in same group and their age-matched controls ( $p1 < 0.001$ ,  $p1 < 0.001$  respectively). As presented in Table 2 Epididymal sperm count, motility and testis weight were significantly lower in rats given Ucn I at doses of 5  $\mu\text{g}$  ( $p1 < 0.05$ ,  $p1 < 0.05$ ,  $p1 < 0.04$  respectively), 10  $\mu\text{g} / \text{kg}$  body weight ( $p1 < 0.006$ ,  $p1 < 0.01$ ,  $p1 < 0.001$  respectively) and 20  $\mu\text{g} / \text{kg}$  body weight ( $p1 < 0.001$ ,  $p1 < 0.001$ ,  $p1 < 0.001$  respectively) compared to their age-matched controls.

## DISCUSSION

Male infertility is a major health problem that represents approximately 30% of all infertilities (Carlsen et al., 1992; Isidori et al., 2006). While stress activates the

hypothalamic–pituitary–adrenal (HPA) axis, it suppresses the hypothalamic–pituitary–gonadal (HPG) axis (Kageyama, 2013, Bhongade et al., 2015). Stress profoundly inhibits the reproductive function by suppressing the pulsatile release of hypothalamic gonadotropin-releasing hormone (GnRH) and consequently luteinizing hormone (LH), at least in part via the corticotrophin-releasing factor (CRF) system as well as through the GABAergic system (Lin et al., 2012, Bhongade et al., 2015).

The release of CRF in response to various stressors suppresses of the HPG axis, especially the GnRH pulse generator in the hypothalamus, and also decreases GnRH mRNA levels via the CRF<sub>1</sub> receptors (Kagayama, 2013). Locally, CRF was found to exert an inhibitory effect on Leydig cell activity in rat testes (Dufau et al., 1993). However, Urocortin I (Ucn I), rather than CRF, is located in rat Leydig cells (Hardy et al., 2005).

Ucn I is an endogenously secreted corticotrophin-releasing factor (CRF)-related peptide. Ucn I is a 40-amino acid peptide that shares 45% homology with CRF (Vaughan et al., 1995; Zhao et al., 1998). In addition, Ucn I exerts its biological activity through CRF<sub>1</sub> and CRF<sub>2</sub> receptors and binds to both types with high affinity (Hsu and Hsueh, 2001). Neurons in the centrally projecting Edinger-Westphal nucleus are the main site of Ucn I synthesis in the mammalian brain, and are assumed to play a role in the stress response. Acute and chronic stress resulted in an increase in Ucn I content of the



Edinger-Westphal nucleus (Derks et al., 2012).

Both CRF and Ucn I contribute to stress responses, cardiovascular and gonadal functions via G protein-coupled seven transmembrane receptors (Vale et al., 1997; Kageyama et al., 1999a; Suda et al., 2004). It is well known that anxiety has a detrimental effect on fertility (Demyttenaere et al., 1988). Ucn I was found to elicit an increase in anxiogenic behavior and potentiate the anxiogenic action of ghrelin as well (Currie et al., 2014).

Therefore, this study is implemented to find out the probable endocrine and reproductive actions of Ucn I in adult male rats via investigation of the effects of exogenous Ucn I on epididymal sperm count, motility, testicular weight and sex hormone profiles. The study of the current hypothesis is that CRF-related peptides act within the gonads, rather than in the periphery, and that their influence on Leydig cells activity is at least partly due to rapid decreases in levels of the steroidogenic enzymes (Herman and Rivier, 2006).

In the present study, Ucn I administration for 42 days significantly decreased the testis weight, epididymal sperm count and motility. These findings are collaborated by the findings of Tao et al. (2007) who hold that Ucn I significantly inhibited the sperm motility and acrosome reaction in a concentration-dependent manner. It inhibited T-type calcium channels in mouse spermatogenic cells, sperm motility and progesterone-evoked sperm acrosome reaction, indicating that inhibition of  $Ca^{2+}$  channels may be a mechanism for the inhibitory effects of Ucn I on male reproductive functions. Ucn I might decrease  $Ca^{2+}$  via inhibiting T-type calcium channels directly in male reproductive cells, instead of binding to its receptors firstly (Tao et al., 2005).

The duration of treatment was documented by Haron et al. (2010). As the germ cells are arranged in specific cell associations, called the stages of the cycle of the seminiferous epithelium. For the rat, the fourteen different stages of each cycle of the seminiferous epithelium last approximately 13 days. The germ cell traverses the different stages of the cycle four times for its complete development, which takes all together 52 days (Clermont, 1972; Karl et al., 1991).

Intraperitoneal administration of Ucn I in the present study significantly decreased the serum testosterone levels. These results are in consistent with findings of Rivier (2008) who reported that intratesticular administration of CRF and Ucn I significantly inhibited the testosterone response to LH-like molecules such as human chorionic gonadotropin (hCG). Ucn I was more effective than CRF in inhibiting Leydig cell responsiveness.

This study further suggested that the intraperitoneal Ucn I administration may be an effective route in inhibition of the release of testosterone from Leydig cells. Ucn I may penetrate the testes after its intraperitoneal injection and acted via a testicular mediated site of action. Blood-borne compounds readily penetrate the

intensely vascularized testes.

In the present study, no significant differences were evident in serum FSH and LH levels between Ucn I-treated rats and their age-matched controls. These results are in agreement with findings of Rivier (2008) who reported that blockade of endogenous LH before Ucn I injection did not alter the inhibitory effect of this peptide regardless of whether it was administered into the general circulation or into the testes.

In addition, it was suggested that the inhibitory influence of CRF-related peptides on testosterone response to gonadotropins is primarily exerted through CRF<sub>1</sub> receptor activation (Li et al., 2005). It is Ucn I, rather than CRF, that is located in rat Leydig cells (Hardy et al., 2005). However, stress-induced suppression of LH pulses was mediated by CRF<sub>2</sub> receptors probably through pituitary Ucn II (Nemoto et al., 2010).

## Conclusion

The results of this study indicate that Ucn I administration significantly reduces testis weight, sperm count, motility and serum testosterone level which does not seem to directly involve the hypothalamic–pituitary–gonadal axis but might be due to a direct effect of Ucn I on testicular tissue and the germ cells.

## Conflict of Interests

The authors have not declared any conflict of interests.

## REFERENCES

- Belsey MA, Moshissi KS, Eliasson R, Paulsen CA, Callegos AJ, Prasad MR (1980). Laboratory manual for the examination of human semen and semen cervical mucus interaction. Press concern.
- Bhongade MB, Prasad S, Jiloha RC, Ray PC, Mohapatra S, Koner BC (2015). Effect of psychological stress on fertility hormones and seminal quality in male partners of infertile couples. *Andrologia* 47(3):336-342.
- Bittencourt JC, Vaughan J, Arias C, Rissman RA, Vale WW, Sawchenko PE (1999). Urocortin expression in rat brain: evidence against a pervasive relationship of urocortin-containing projections with targets bearing type 2 CRF receptors. *J. Comp. Neurol.* 415:285-312.
- Carlsen E, Giwercman A, Keiding N, Skakkebaek NE (1992). Evidence for decreasing quality of semen during past 50 years. *BMJ* 305:609-613.
- Clermont Y (1972). Kinetics of spermatogenesis in mammals: seminiferous epithelium cycle and spermatogonial renewal. *Physiol. Rev.* 52:198-235.
- Currie PJ, Schuette LM, Wauson SE, Voss WN, Angeles MJ (2014). Activation of urocortin 1 and ghrelin signaling in the basolateral amygdala induces anxiogenesis. *Neuroreport* 25(1):60-64.
- Demyttenaere K, Nijs P, Steeno O, Koninckx PR, Everse-Kiebooms G (1988). Anxiety and conception rates in donor in semi nation. *J. Psychosom. Obstet Gynaecol.* 8:175-181.
- Derks NM, Pintér O, Zuure W, Ledent C, Watanabe M, Molnár CS, Wei Y, Roubos EW, Wu S, Hrabovszky E, Zelena D, Kozicz T (2012). Cannabinoid modulation of midbrain urocortin 1 neurones during

- acute and chronic stress. *J. Neuroendocrinol.* 24(11):1447-1461.
- Dufau ML, Tinajero JC, Fabbri A (1993). Corticotropin-releasing factor: an antireproductive hormone of the testis. *FASEB J* 7:299-307.
- Fekete ÉM, Zorrilla EP (2007). Physiology, pharmacology and therapeutic relevance of urocortins in mammals: Ancient CRF paralogs. *Front. Neuroendocrinol.* 28:1-27.
- Hardy MP, Gao HB, Dong Q, Ge R, Wang Q, Chai WR, Feng X, Sottas C (2005). Stress hormone and male reproductive function. *Cell Tissue Res.* 322:147-153.
- Haron MN, D'Souza UJ, Jaafar H, Zakaria R, Singh HJ (2010). Exogenous leptin administration decreases sperm count and increases the fraction of abnormal sperm in adult rats. *Fertil Steril.* 93(1):322-324.
- Herman M, Rivier C (2006). Activation of a neural brain-testicular pathway rapidly lowers Leydig cell levels of the steroidogenic acute regulatory protein and the peripheral-type benzodiazepine receptor while increasing levels of neuronal nitric oxide synthase. *Endocrinology* 147(1):624-633.
- Hsu SY, Hsueh AJ (2001). Human stresscopin and stresscopin-related peptide are selective ligands for the type 2 corticotropin releasing hormone receptor. *Nat. Med.* 7:605-611.
- Idris MH, Budin SB, Osman M, Mohamed J (2012). Protective role of hibiscus sabdariffa calyx extract against streptozotocin induced sperm damage in diabetic rats. *EXCLI J.* 11:659-669.
- Isidori AM, Pozza C, Gianfrilli D, Isidori A (2006). Medical treatment to improve sperm quality. *J. Reprod. Biomed.* 12:704-714.
- Kageyama K (2013). Regulation of gonadotropins by corticotropin-releasing factor and urocortin. *Front. Endocrinol.* 4:12.
- Kageyama K, Bradbury MJ, Zhao L, Blount AL, Vale WW (1999a). Urocortin messenger ribonucleic acid: tissue distribution in the rat and regulation in thymus by lipopolysaccharide and glucocorticoids. *Endocrinology* 140(12):5651-5658.
- Karl PI, Katz R, Daum F, Fisher SE (1991) Mercaptopurine and spermatogenesis in the young rat. *Dig. Dis. Sci.* 36:1569-1573.
- Karteris E, Hillhouse EV, Grammatopoulos D (2004). Urocortin II is expressed in human pregnant myometrial cells and regulates myosin light chain phosphorylation: potential role of the type-2 corticotropin-releasing hormone receptor in the control of myometrial contractility. *Endocrinology* 145:890-900.
- Khaki A, Nouri M, Fathiazad F, Ahmadi-Ashtiani HR, Rastgar H, Rezazadeh. (2009). Protective Effects of Quercetin on Spermatogenesis in Streptozotocin induced Diabetic Rat. *J. Med. Plant* 9(35):143-150.
- Kihara N, Fujimura M, Yamamoto I, Itoh E, Inui A, Fujimiya M (2001). Effects of central and peripheral urocortin on fed and fasted gastroduodenal motor activity in conscious rats. *Am. J. Physiol. Gastrointest. Liver Physiol.* 280:G406-G419.
- Lee S, Braden B, Kang SS, Rivier C (2011). Urocortins are Present in the Rat Testis. *Neuropeptides* 45(2):131-137.
- Lewis K, Li C, Perrin MH, Blount A, Kunitake K, Donaldson C, Vaughan J, Reyes TM, Gulyas J, Fischer W, Bilezikjian L, Rivier J, Sawchenko PE, Vale WW (2001). Identification of urocortin III, an additional member of the corticotropin-releasing factor (CRF) family with high affinity for the CRF2 receptor. *Proc. Natl. Acad. Sci USA* 98:7570-7575.
- Li XF, Bowe JE, Lightman SL, O'Byrne KT (2005). Role of corticotropin-releasing factor receptor-2 in stress-induced suppression of pulsatile luteinizing hormone secretion in the rat. *Endocrinology* 146(1):318-322.
- Lin YS, Li XF, Shao B, Hu MH, Goundry AL, Jeyaram A, Lightman SL, O'Byrne KT (2012). The Role of GABAergic Signalling in Stress-Induced Suppression of Gonadotrophin-Releasing Hormone Pulse Generator Frequency in Female Rats. *J. Neuroendocrinol.* 24(3):477-488.
- Nemoto T, Iwasaki-Sekino A, Yamauchi N, Shibasaki T (2010). Role of urocortin 2 secreted by the pituitary in the stress-induced suppression of luteinizing hormone secretion in rats. *Am J Physiol Endocrinol Metab.* 299(4):E567-E575.
- Oki Y, Sasano H (2004). Localization and physiological roles of urocortin. *Peptides* 25:1745-1749.
- Petraglia F, Florio P, Benedetto C, Marozio L, Di Blasio AM, Ticconi C (1999). Urocortin stimulates placental adrenocorticotropin and prostaglandin release and myometrial contractility *in vitro*. *J. Clin. Endocrinol. Metab.* 84:1420-1423.
- Rebar RW, Morandini IC, Petze JE, Erickson GF (1982). Hormonal basis of reproductive defects in athymic mice: reduced gonadotropins and testosterone in males. *Biol. Reprod.* 5:1267-1276.
- Rivier CL (2008). Urocortin 1 Inhibits Rat Leydig Cell Function. *Endocrinology* 149(12):6425-6432.
- Rodríguez B, Bermudez L, Poncede Leon E, Castro L (1983). Presented at the third world congress of behavior therapy. Washington DC; The relationship between infertility and anxiety: some preliminary findings. December 8–11.
- Suda T, Kageyama K, Sakihara S, Nigawara T (2004). Review Physiological roles of urocortins, human homologues of fish urotensin I, and their receptors. *Peptides* 25(10):1689-701.
- Tao J, Min Lin, Jiahao Sha, Gregory Tan, Tuck Wah Soong, Shengnan Li (2007). Separate Locations of Urocortin and its Receptors in Mouse Testis: Function in Male Reproduction and the Relevant Mechanisms. *Cell. Physiol. Cell Physiol. Biochem.* 19:303-312.
- Tao J, Wu Y, Chen J, Zhu H, Li S (2005). Effects of urocortin on T-type calcium currents in mouse spermatogenic cells. *Biochem. Biophys. Res Commun.* 329:743-748.
- Tietz NW (1995). *Clinical Guide to Laboratory Tests*, 3rd Ed., W.B. Saunders Company, Philadelphia pp. 509-580.
- Vale W, Vaughan J, Perrin MH (1997). Corticotropin-releasing factor (CRF) family ligands and their receptors. *Endocrinologist* 7:3S-9S.
- Vaughan J, Donaldson C, Bittencourt J, Perrin MH, Lewis K, Sutton S, Chan R, Turnbull AV, Lovejoy D, Rivier C, Rivier J (1995). Urocortin, a mammalian neuropeptide related to fish urotensin I and to corticotropin-releasing factor. *Nature* 378(6554):287-292.
- Yutaka O, Hironobu S (2004). Localization and physiological roles of urocortin. *Peptides* 25:1745-1749.
- Zhao L, Donaldson CJ, Smith GW, Vale WW (1998). The structures of the mouse and human urocortin genes (Ucn and UCN). *Genomics* 50:23-33.



# African Journal of Pharmacy and Pharmacology

## Related Journals Published by Academic Journals

- *Journal of Medicinal Plant Research*
- *African Journal of Pharmacy and Pharmacology*
- *Journal of Dentistry and Oral Hygiene*
- *International Journal of Nursing and Midwifery*
- *Journal of Parasitology and Vector Biology*
- *Journal of Pharmacognosy and Phytotherapy*
- *Journal of Toxicology and Environmental Health Sciences*



**academicJournals**